## Oxidants and Antioxidants in COPD Associated with Tobacco Smoke and Biomass Exposure

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## ABSTRACT

## BACKGROUND

Chronic Obstructive Pulmonary Disease (COPD) is one of the leading causes of chronic morbidity and mortality. Apart from tobacco smoke, biomass fuel has been implicated as an important etiological factor for development of COPD. Oxidantantioxidant imbalance is known to play a key role in pathophysiology of COPD. The study was undertaken to evaluate the role of oxidative stress and antioxidant status among COPD cases due to tobacco smoking and biomass exposure.

## METHODS

Serum MDA, and erythrocyte SOD and GSH levels, were estimated among 40 COPD cases due to tobacco smoking (Group 1), 20 COPD cases due to biomass exposure (Group 2). 40 age and sex matched healthy controls (Group 0) were also included. Serum MDA, SOD and GSH were measured calorimetrically by TBA method, Marklund & Marklund method and method by Beutler et al respectively.

## RESULTS

IBM SPSS Ver. 20 was used for statistical analysis and preparation of tables. Significantly higher levels of MDA were seen among COPD cases due to tobacco smoke ( $58.08\pm52$  vs  $15.3705\pm6.6$ ; p value <0.01) compared to controls. SOD levels were significantly lower in both case groups compared to controls ( $1123.3\pm301.2$ ,  $1147.01\pm200.5$  vs  $1315.23\pm209.1$ ; p value<0.01). GSH levels were lower in tobacco smoking group when compared to biomass exposed group ( $7.98\pm2.7$  vs  $9.61\pm2.1$ ; p value 0.01). Positive correlation was found between FEV1% and SOD in group 1 cases.

## CONCLUSIONS

The results support the hypothesis of presence of increased oxidative stress and oxidant-antioxidant imbalance in pathogenesis of COPD. It plays an important role in disease severity which is higher among COPD in tobacco smokers compared to biomass exposed COPD.

## **KEY WORDS**

COPD, Smoking, Biomass, Oxidants, Antioxidants

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## BACKGROUND

Aetiology of COPD in India: Chronic obstructive pulmonary disease (COPD), a common preventable and treatable disease, is characterized by persistent airflow limitation that is usually progressive and is associated with an enhanced chronic inflammatory response in the airways and the lung to noxious particles or gases.<sup>[1]</sup> Exacerbations and co morbidities contribute to the overall severity in individual patients. Much of the increase in incidence of COPD is associated with projected increase in tobacco use and the indoor exposure to smoke from the combustion of solid biomass fuel, for heating and cooking.<sup>[2]</sup> Significantly ratios for COPD in India for both Male: Female and smoker: nonsmoker are not as high as in the Western populations. This is largely attributed to the indoor air pollution from domestic combustion of solid biomass fuels.[3],[4],[5] Commonly used solid biomass fuels for cooking are cow dung cake, wood and coal. Environmental tobacco smoke (ETS) from passive smoking mainly from male smokers in the house is also another important risk factor for COPD in non-smoker women.<sup>[3]</sup>

Cigarette smoke and inflammation: Oxidative stress has been attributed to play the central role in the pathogenesis of COPD. In addition to causing direct injury to the respiratory tract, oxidative stress triggers and exacerbates chiefly three other mechanisms namely inflammation, apoptosis and protease-antiprotease imbalance.<sup>[6],[7]</sup> Cigarette smoke contains approximately five thousand toxic compounds, including potent oxidants (Approximately 1014 free radicals per inhalation) such as acrolein, hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hydroxyl (OH.) and organic free radicals.<sup>[8]</sup> Reactive oxygen species (ROS) directly recruit inflammatory cells such as neutrophils, eosinophils, lymphocytes and macrophages resulting in inflammation.<sup>[9]</sup> ROS cause overexpression of the genes of proinflammatory mediators (e.g. tumour necrosis factor [TNF]- $\alpha$ , interleukin[IL]-1, and interleukin[IL]-8) via transcription factor NF-kb(Nuclear factor kappa B) and AP-1 (Activator protein-1), thus further recruiting inflammatory cells.[10] ROS can also form reactive aldehydes by lipid peroxidation of membrane phospholipids like 4-hydroxy-2nonenal (4HNE) and MDA (Malondialdehyde) which are capable of inducing caspase (A major promoter of cell apoptosis).[11]

Human biological antioxidants: In normal cell, there is an appropriate pro-oxidant: antioxidant balance. Indigenous compounds and reactions disposing of these species, scavenging them, suppressing their formation or opposing their actions are antioxidants and include compounds such as NADPH (Reduced nicotinamide adenine dinucleotide phosphate), GSH (reduced glutathione), ascorbic acid, vitamin E, superoxide dismutase (SOD), glutathione peroxidase (GPx) etc.<sup>[12]</sup> SOD and GSH associated enzymes are enzymatic antioxidants which are active at the beginning of reaction through which reactive species are formed, and this avoids accumulation of  $O_2$ - and  $H_2O_2$ .<sup>[13]</sup> There are three isomers of SOD, all neutralize superoxide. GSH is one of the primary nonenzymatic antioxidants in lung and exists in epithelial lining fluid, reduces H<sub>2</sub>O<sub>2</sub> and lipid peroxides and neutralizes xenobiotic radicals.<sup>[14]</sup>

Effects of oxidative stress: Oxidative stress causes peroxidation of Polyunsaturated Fatty Acid (PUFA) present in cell membrane phospholipids. This leads to alteration in the structure and permeability of the cell membrane, resulting in loss of ion exchange selectivity, release of contents of organelles, i.e. hydrolytic enzymes of lysosome and formation of cytotoxic products such as MDA.<sup>[15]</sup>

#### **Measurements of Human Oxi-Antioxidants**

Oxidative stress can be measured through indirect quantification of products of lipid peroxidation like MDA, in the alveolar space, in the exhaled breath condensate, in the sputum and the blood.<sup>[16]</sup> Simultaneous estimation of antioxidant status can be made by measurement of serum levels of antioxidants like SOD, GSH.

## METHODS

Place of study and patient selection: A total of 100 subjects including cases and controls had participated in the study. 60 stable COPD cases, none having acute exacerbation at the time of study, diagnosed clinically and confirmed with spirometry attending or admitted in Department of Pulmonary and Chest Medicine, Calcutta National Medical College and Hospital, Kolkata, West Bengal. Sample size was taken based on the convenience of the study. Approval from the institutional ethical committee was obtained for the study. The cases were divided into two groups based on current smoking history and history of exposure to biomass consisting of 40 (Group 1) and 20 (Group 2) cases respectively. Similarly, 40 age and sex matched healthy controls without history of smoking or biomass exposure were selected from persons who accompanied the patients and also from health care workers who participated voluntarily in the study (Group 0). Subjects with dual exposure to tobacco smoke and biomass were excluded from this study. Informed consent was obtained from all the subjects prior to the study.

Subject exclusion criteria and analyte estimation: Patients with pleural disease, malignancy of lung, tuberculosis, human immunodeficiency virus (HIV) infection, diffuse parenchymal lung disease (DPLD), diabetes mellitus, dyslipidemia, hypertension, connective tissue disorders, chronic renal or hepatic diseases were excluded from the study. Chest skiagram PA view, pulmonary function test (PFT) with bronchodilator reversibility were done in all cases and controls. About 6 ml of blood was drawn from a large peripheral vein under aseptic precaution after overnight fasting with a heparinised syringe and was transported immediately to the laboratory, Department of Biochemistry, Calcutta National Medical College and Hospital. The sample was then centrifuged to separate serum and prepare RBC lysate. Serum MDA activity was measured in serum by thiobarbituric acid method.[17] SOD and reduced GSH activities were measured in RBC lysate by methods of Marklund and Marklund<sup>[18]</sup> and Beutler et al<sup>[19]</sup> respectively.

## Statistical Analysis

Statistical analysis was done with IBM SPSS Ver. 20. Chisquare test was performed to analyze the age and sex distribution pattern among cases and controls. Post-hoc ANOVA with and without Bonferroni correction was performed to estimate the statistical significance between variables like MDA, SOD and GSH among case groups and controls.

## RESULTS

Age comparison among case groups and controls: The mean age distribution among COPD in smokers, COPD among biomass exposed and controls, selected for the study were close to each other as much as possible (Table 1) [Chi-square Test: p-value >0.068, degree of freedom=1].

Comparison of MDA, SOD and GSH levels among case groups and controls: The mean  $\pm$  SD values for MDA, SOD, and GSH among the case groups (1 and 2) and controls (0) were represented in (Table 2). The values of MDA were significantly higher in Group 1 cases when compared to Group 0 (p-value < 0.001). No statistical difference was seen between cases and between Group 2 and Group 0. SOD levels were significantly lower among the case groups when compared to controls (p value < 0.05). Higher level of GSH was found in biomass exposed COPD cases than COPD with smoking history (p value < 0.05) (Table 3A, Table 3B).

Correlation between FEV1% and various oxiantioxidants: In Group 1, FEV1% was positively correlated to SOD value (p value <0.015). SOD and GSH values were also correlated to each other among smokers (Table 4). No similar correlation existed for FEV1%, MDA, SOD and GSH among the group 2 cases and controls.

Groups		Group 1 (n=40)	Group 2 (n=20)		Group 0 (n=40)
Age (in yrs.) Mean ± SD		59.15±8.92	58.6±9.43		55±5.16
Sex	Male	32	4		31
	Female	8	16		9
Smoking History (Pack yrs)		27	Nil		Nil
Biomass Exposure (yrs)		Nil	32		Nil
FEV1 (ml)		845	753.75	1373.875	
FVC (ml)		1611	1373.87		
FEV1/ FVC %		57	55.95	44.483193	
FEV1% of predicted value		44.1	44.48		
GOLD stage of airflow limitation			3		
Table 1. Distribution Pattern for Age, Sex, Smoking History, Biomass Exposure and COPD Levels among Cases and Controls					

6		MDA	SOD	GSH
Groups		(nmol/ml)	(units/gm of Hb)	(µmol/gm of Hb)
Smokers	Mean	58.0892 ±	1123.3922 ±	7.9755 ±
(Group 1: n=40)	±S.D.	52.53836	301.24220	2.70204
Biomass	Mean	35.8219 ±	1147.0161 ±	9.6113 ±
(Group 2:n=20)	±S.D.	23.73494	200.58413	2.09725
Control	Mean	15.3705 ±	1315.2305 ±	8.2895 ±
(Group 0: n=40)	±S.D.	6.67772	209.16684	1.08898

Table 2. Mean Serum Levels for MDA, SOD and GSH among Cases and Controls

F-test of ANOVA	Sum of Squares	df	<b>Mean Square</b>	F	Sig.
Between Groups	39760814.19	2	19880407.099	699.36	.000
Within Groups	4349244.795	153	28426.437		
Total	44110058.99	155			
Table 3A. Comparison Between and within Case Groups and Controls					

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Multiple Co	Multiple Comparisons Post-hoc ANOVA with Bonferroni Correction				
Dependent Variable	(X) Grouping	(Y) Grouping	Mean Difference (X-Y)	Std. Error	Level of Significance (p Value)
	0 vs	1	- 42.71867*	7.86790	.000
MDA	0 vs	2	- 20.45145	9.63617	.109
	1 vs	2	22.26722	9.63617	.069
	0 vs	1	191.83835*	55.65845	.003
SOD	0 vs	2	168.21445*	68.16740	.046
	1 vs	2	-23.62390	68.16740	1.000
	0 vs	1	.31405	.46227	1.000
GSH	0 vs	2	-1.32180	.56616	.065
	1 vs	2	-1.63585*	.56616	.014
Tab	Table 3B. Comparison for MDA, SOD and GSH among				
case Groups and Controls					
Comparison of different oxidative stress markers in three study groups					
The mean difference is significant at the <0.05 level					
Control group=0(X), Smokers with COPD=group1(Y), Biomass exposed					
COPD=group2(Y)					
(X-Y): Differen	(X-Y): Difference between mean values between cases and controls and those				
between the ty	between the two case groups. Abbreviation- vs: Versus				

Parameters	r Value	p Value
FEV1%: MDA	061	.711
FEV1%: SOD	.383	.015
FEV1%: GSH	.148	.368
MDA: SOD	.023	.886
MDA: GSH	181	.263
SOD: GSH	.356	.024

Table 4. Pearson's Correlation Between MDA, SOD, GSH and FEV1% in COPD Cases with History of Smoking Tobacco





## DISCUSSION

Serum MDA levels alteration among smokers and biomass exposed subjects: It is known that oxidative stress is a major pathogenic component of airway inflammation that is characteristic of COPD.<sup>[20]</sup> Oxidative radicals cause damage to cellular components including membrane lipids, protein, carbohydrate and DNA.[21] It is well known that both cigarette and biomass exposure causes COPD in which free radicals and ROS increase.<sup>[22],[23]</sup> In this study, we found that plasma MDA level, an indicator of lipid peroxidation was higher in both smoking and biomass exposed COPD compared to healthy controls. PUFA and Fatty acids are major targets for free radical attack, resulting in lipid peroxidation, that continues as a chain reaction.<sup>[24]</sup> It had been shown that the levels of MDA, produced due to lipid peroxidation reaction in plasma are correlated inversely with the FEV1 percent in a population study.<sup>[25]</sup> There are reports indicating increased MDA level in both cigarette smoking and biomass exposure.<sup>[25]</sup> It is reported that there is significant decrease in antioxidant enzyme activity in females exposed to biomass fuel.

## **Depleted SOD Stores in COPD**

SOD functions as a scavenger of O<sub>2</sub>. Radical in the body. The level of SOD is decreased in oxidative stress which plays an important role in pathogenesis of various diseases including COPD.<sup>[26]</sup> Studies of Raghunath R Rai et al,<sup>[26]</sup>Nagaraj et al<sup>[27]</sup> and Ahmad A et al<sup>[28]</sup> showed decreased level of SOD amongst patients with COPD. This is in accordance to our study results. However, Montano et al<sup>[25]</sup> have found increased SOD levels in COPD patients compared to healthy controls. These alterations in antioxidant enzymes such as SOD emphasize to redox imbalance in COPD patients. Mechanism involved in variable SOD activity is due to increased production of free radicals in COPD patients resulting in increased SOD biosynthesis as a protective mechanism as well as its increased consumption, leading to depleted intracellular levels.

#### **GSH Level Variability among Cases**

Under non-stress condition, most of the intracellular glutathione is stored in reduced form (GSH). During increased oxidative stress by tobacco smoke or biomass, the free sulfhydryl (-SH) groups become oxidized resulting in loss of GSH. The gaseous phase of cigarette smoke irreversibly reacts with GSH to form GSH derivatives which cannot be reduced back, thus depleting the total GSH pool.<sup>[29]</sup> In our study, GSH among biomass exposed COPD was statistically higher than tobacco smokers with COPD (p value <0.02). Toth and colleagues (1986) stated that RBC glutathione plays a prominent role in detoxification of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>). Increased GSH in RBC of COPD patients is presumably due to constant exposure to oxidative stress, resulting in induction of protein synthesis.<sup>[30]</sup> However the activities of glutathione synthesis and redox system enzymes like glutathione peroxidase (GPx) and glucose-6-phosphate dehydrogenase (G6PD), gamma-glutamyl cysteine synthetase are transiently decreased in alveolar epithelial cells after chronic exposure to cigarette smoke condensate. This is due to action of highly electrophilic free radicals on the active sites of these enzymes, thus there is a time-dependent depletion of intracellular soluble GSH.<sup>[31]</sup> Studies had also shown a gradual age dependent decrease in serum glutathione levels among smokers.<sup>[32]</sup>

#### High Pathogenicity of Tobacco Smoke in Causing COPD

Studies have shown tobacco smoke induced COPD was associated with worse emphysema index in HRCT compared to biomass exposed COPD. This suggests tobacco smoke is more aggressive and leads to more parenchymal destruction than other forms of inhaled toxins.<sup>[32]</sup> Much in the same way our study explains lower GSH levels in tobacco smoke induced COPD cases compared to biomass exposed COPD cases.

# Exposure to Both Tobacco Smoke and Biofuels Cause COPD

These results indicate the role of oxidative stress in causing COPD associated with tobacco smoke and inhalation of toxin for multiple hours per day, during use of solid biomass fuel for cooking food, a trend and a job done mostly by females all across India.<sup>[33]</sup> Since unlike contents of tobacco smoke the details of various chemicals present in commonly used biofuels in rural India are yet not analyzed their individual mechanism as oxidants in pathogenesis of COPD requires further investigation.

## CONCLUSIONS

Our study confirmed the presence of an oxidant-antioxidant imbalance in COPD subjects supporting the concept of systemic oxidative stress in this condition.

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