ANALYSIS OF ABO GROUP DISCREPANCIES IN A TERTIARY CARE CENTRE SOUTH INDIA

Bonagiri Shanthi1, K. Ravi Babu2

1Associate Professor, Department of IHBT, Nizam’s Institute of Medical Sciences.
2Senior Resident, Department of Bio-Chemistry, Nizam’s Institute of Medical Sciences.

ABSTRACT

BACKGROUND
ABO system remains the most important blood group system in transfusion and organ transplantation medicine. ABO typing and ABO incompatibility testing remain the foundation of all pretransfusion testing. An inverse reciprocal relationship exists between the presence of A and B antigens on red cells and the presence of Anti-A and Anti-B or both in sera. Discrepancies in ABO grouping can be characterised by inappropriate results in either cell or serum grouping.

The aim of this study is to analyse commonly occurring ABO group discrepancies in order to standardise resolving procedure and avoid delay in pretransfusion testing.

MATERIALS AND METHODS
Descriptive study period included from 2012 to 2016, the pretransfusion tests done by tube and gel column agglutination method. Total 28,024 blood grouping and typing tests were retrospectively evaluated in the study.

RESULTS
ABO blood group discrepancies were identified in 1331 cases, which is equal to 4.75% of total tests. Subgroups of A in 1089 cases (81%), Autoantibodies in 187 cases (14.5%), Alloantibodies in 33 cases (2.56%), 16 cases were typed as Bombay phenotype and 8 as H deficient probably, Para-Bombay phenotype and two cases of multiple myeloma and heparin contaminated sample causing discrepancy in cell grouping. These cases were identified during pretransfusion workup.

CONCLUSION
Resolving ABO and Rh grouping and typing discrepancy is crucial in pretransfusion testing and selecting proper blood and blood products in transfusion services. Most of the problems can be resolved by basic immunohaematological techniques.

KEYWORDS
ABO Blood Group System, Discrepancies, Red Cell Serology.

reagent-dependent antibody or rouleaux, transplantation, acquired A antigen, acquired B-like antigen and out-of-group transfusion; iii) Mixed-field RBC reactivity for recent transfusion, transplantation, foeto-maternal haemorrhage and chimerism; iv) Weak/missing serum reactivity due to age (< 4 – 6 months old or elderly persons), ABO subgroups, hypogammaglobulinaemia and transplantation; v) Extra serum reactivity caused by cold auto- or allo-antibodies, antibodies to reagent constituent, excess serum protein, transfusion of plasma, transplantation or infusion of intravenous immunoglobulin.[5]

**Aim of the Study**
To analyse commonly occurring ABO group discrepancies in order to standardise resolving procedure and avoid delay in pretransfusion testing.

**MATERIALS AND METHODS**
This is a descriptive study over a period of 4 years (2012 - 2016). Total of 28,024 ABO blood grouping tests were analysed retrospectively for discrepancies at the Department of Transfusion Medicine, referral centre. ABO typing was done using gel column agglutination method. The ABO-Rh D/Reverse Typing Cassette consists of six columns, five of which contain gel, buffers and reagents including anti-A, anti-B, anti-D, A1 cells and B cells. The sixth column serves as a negative control well; it only contains the red cell suspension of the sample. During the centrifugation of the cassettes, non-agglutinated red cells sink to the bottom of the column, while agglutinated red cells remain at different levels of the column according to the amount of agglutination. The reactions are graded as Traces 1+, 2+, 3+ or 4+ to represent the strength of the reaction.

Antibodies were screened by column agglutination technology (CAT) using commercially available three cell antigen panel by Coombs gel card. Whenever antibody screening was positive, extended eleven cell panel was used for antibody identification using low ionic strength saline (LISS).

In ABO typing by the reference manual method for forward typing, one drop each of anti-A and anti-B reagent was added to each test tube containing one drop of 2% - 5% red cell suspension. For reverse typing, one drop of each of A and B cell reagents was added to each test tube containing two drops of the plasma sample. As an auto control, the red cells and serum of the sample were reacted together in another test tube. Samples that showed discrepant results after retesting were tested by additional manual methods were re-evaluated by using anti-A, anti-H and anti-A1 (Dolichos biflorus and Ulex europaeus lecin) reagents.[6] If cold agglutinins were suspected, the sample was collected and immediately incubated at 37°C and at room temperature for 30 mins. after saline washing and then retested. Interpretation of test results were described as the reaction grades obtained with each method. Finally, results showing extra cell or serum reactions (including any reaction grade) were regarded as discrepant results.

**Statistical Analysis**
The percentage of discrepant results was calculated.

**RESULTS**
Discrepancy was found in 1331 (4.75%) of 28,024 ABO grouping tests. Total sub-groups of A and B detected with Anti-A1, AB and H lectin were 1089 out of 28,024 blood grouping and typing tests accounting for 81% (Fig. 1) of blood group discrepancies in our lab, labelled as A2 or A2B, A3 and Bx. Discrepancy caused by red cell autoantibodies was observed in 187 cases (14.5%), in that causing panagglutination was observed in 69/187 cases by cold autoantibodies (37%), rest of discrepancies were because of mixed and warm autoantibodies. Rare blood group phenotypes, which caused unexpected antibody considered as Bombay in 16 patients (1.2% of discrepancies) and Para-Bombay in 6 patients. Red cell alloantibodies accounted for 2.5% of total ABO discrepancies. Three and 11 cell panel antibody identification revealed red cell antibodies like Anti-D, Anti-E, Kell, Lewis and MNS. Forward group discrepancy was observed in multiple myeloma[7] and patient on heparin therapy. Mixed field reaction was observed in one case of allogenic bone marrow stem cell transplantation and group O red cell transfusions to A/B/AB patients. The discrepant results were grouped into IV types (Table 2).

**Table 1. Types of ABO Group Discrepancies and their Serological Reactions**

<table>
<thead>
<tr>
<th>Discrepancy</th>
<th>Diagnosis</th>
<th>Bombay H-Absent</th>
<th>A2B</th>
<th>Bone Marrow Tx</th>
<th>Old Age</th>
<th>Cold Auto-antibody</th>
<th>Alloantibody</th>
<th>H-Deficient</th>
<th>Bx</th>
<th>H-Deficient</th>
<th>Warm Auto-Antibody</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group of Discrepancy</td>
<td>IV</td>
<td>II</td>
<td>I</td>
<td>I</td>
<td>I</td>
<td>IV</td>
<td>IV</td>
<td>II</td>
<td>II</td>
<td>II &amp; IV</td>
<td>IV</td>
</tr>
<tr>
<td>Type of Discrepancy</td>
<td>Extra antibody</td>
<td>Weak Anti A expression</td>
<td>Mixed field reaction</td>
<td>Missing antibody</td>
<td>Extra antibody</td>
<td>Extra antibody</td>
<td>No antigen expression</td>
<td>Weak Anti B expression</td>
<td>Extra antibody</td>
<td>Extra antibody</td>
<td></td>
</tr>
<tr>
<td>Serum</td>
<td>Unknown</td>
<td>B</td>
<td>B</td>
<td>AB</td>
<td>O</td>
<td>O</td>
<td>B</td>
<td>B</td>
<td>Unknown</td>
<td>O</td>
<td></td>
</tr>
<tr>
<td>Cell</td>
<td>O cells</td>
<td>4+</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>2+</td>
<td>3+</td>
<td>0</td>
<td>0</td>
<td>4+</td>
<td>2+</td>
</tr>
<tr>
<td>B cells</td>
<td>4+</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>3+</td>
<td>3+</td>
<td>2+</td>
<td>0</td>
<td>4+</td>
<td>4+</td>
<td>4+</td>
</tr>
<tr>
<td>A1 cells</td>
<td>4+</td>
<td>1+</td>
<td>4+</td>
<td>0</td>
<td>4+</td>
<td>4+</td>
<td>0</td>
<td>4+</td>
<td>4+</td>
<td>4+</td>
<td>4+</td>
</tr>
<tr>
<td>Cell Grouping</td>
<td>Anti-H</td>
<td>0</td>
<td>3+</td>
<td>3+</td>
<td>2+</td>
<td>2+</td>
<td>2+</td>
<td>0</td>
<td>4+</td>
<td>Weak and delayed expression</td>
<td>2+</td>
</tr>
<tr>
<td>Anti-B</td>
<td>1+</td>
<td>2+</td>
<td>0</td>
<td>2+</td>
<td>2+</td>
<td>0</td>
<td>4+</td>
<td>4+</td>
<td>2+</td>
<td>2+</td>
<td></td>
</tr>
<tr>
<td>Anti-A1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>2+</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>4+</td>
<td>4+</td>
<td>4+</td>
</tr>
<tr>
<td>Anti-D</td>
<td>0</td>
<td>4+</td>
<td>4+</td>
<td>4+</td>
<td>4+</td>
<td>0</td>
<td>0</td>
<td>4+</td>
<td>4+</td>
<td>4+</td>
<td>4+</td>
</tr>
<tr>
<td>Anti-B</td>
<td>0</td>
<td>3+</td>
<td>MF</td>
<td>3+</td>
<td>4+</td>
<td>4+</td>
<td>0</td>
<td>0</td>
<td>2+</td>
<td>2+</td>
<td></td>
</tr>
<tr>
<td>Anti-A</td>
<td>0</td>
<td>2+</td>
<td>0</td>
<td>2+</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Sl. No.</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td>4</td>
<td>5</td>
<td>6</td>
<td>7</td>
<td>8</td>
<td>9</td>
<td>10</td>
<td></td>
</tr>
</tbody>
</table>
DISCUSSION

ABO blood group testing is the basis for pretransfusion testing to select accurate blood unit for transfusion. In addition, ABO blood group is very important in identification, blood donor group, bone marrow and solid organ transplantation. An ABO incompatible red blood cells transfusion is a leading cause of death from transfusion.[3] Subgroups, alloantibodies and autoantibodies are the common reasons for ABO discrepancies in routine immunohaematology practice.[4,5]

In our descriptive study, after sub-groups autoantibodies are the main reason for ABO group discrepancies. And it is essential to decide ABO group in such cases to select the least incompatible blood unit with limited inventory of O, RH negative red cells. For that we can use auto-adsorption of antibodies at cold/warm temperature and elution technique to resolve the discrepancy. Cold agglutinins usually cause panagglutination and cell group shows AB group and serum O group. In such cases, instructions were given to collect blood sample in warm condition (37°C).[6] Auto control test is useful in differentiating warm autoagglutinins and clinically significant alloantibodies if there is an unexpected antibody in serum grouping.

In this study ABO discrepancies occurred because of alloantibodies at all phases of reactions by different types of antibodies, mainly IgM at 4°C and room temperature (Anti-M and Anti-E) at 37°C and AHG phase Anti-D, Anti-C, Kidd etc. Antibody identification with 3 cell and 11 cell panel was done at 3 phases to detect antibody, resolve serum discrepancy and selection of suitable blood unit.[9]

Identification of sub-groups is possible with lectins Anti-A1, Anti-H and Anti-AB reagents. In our study, these reagents in routine red cell serology and saliva testing detected subgroups, Bombay phenotype and Para-Bombay phenotypes. Family ABO group testing helps in confirming H-deficient/Bombay phenotype and other rare blood groups.[10] Blood group genotyping confirms the diagnosis for such discrepant results.

Thirty nine year old female diagnosed case of Acute Myeloid Leukaemia, O RH positive group received allogenic peripheral blood stem cell transplantation from stem cell donor of B RH positive group, after three months ABO grouping and typing showed mixed field reaction with Anti-B in cell group. Blood group chimerism is an intrinsic characteristic of ABO non-identical HPC transplantation. The erythrocyte chimerism[11] helps in using the blood products with donor ABO typing or not. In our case, patient showed full-donor chimera indicating successful engraftment.

CONCLUSION

ABO blood grouping is the first and foremost investigation in blood transfusion services. If ABO discrepancies are found, they have to be resolved by starting with the basic direct Coomb's test, Indirect Coomb's test and auto control. Most of them can be resolved serologically even before resorting for high end investigations. However, it is essential to strictly adhere to the standard operating procedures, from sample collection to laboratory practices, to avoid delayed turnaround time for issuing blood components. In case of delay in resolving the discrepancy coordination between the clinician, laboratory values, ongoing treatment and transfusion medicine specialist are mandatory to decide the blood product to be selected for transfusion.

Molecular ABO genotyping[12] in confirming rare subgroups, Bombay/Para-Bombay phenotypes is very useful to reaffirm our basic red cell serology methods to resolve these unusual ABO discrepancies.

REFERENCES
