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AN ANNOTATED CHECKLIST OF MOSQUITO FAUNA WITH VECTOR BIONOMICS IN NILGIRI HILLS, SOUTHERN INDIA

P. J. Bhuyan, J. Hiriyan, P. Chandrasekaran, C. Annadurai

1. Public Health Specialist. National Centre for Disease Control, Coonoor.
2. Senior Technical Officer, CRME, Madurai.
3. Technical Staff, National Centre for Disease Control, Madurai.
4. Technical Staff, National Centre for Disease Control, Madurai.

CORRESPONDING AUTHOR:

P. J. Bhuyan,
National Centre for Disease Control, Coonoor Branch,
Shanmugapriya, Brook Lands.
Coonoor – 643101, The Nilgiris, Tamil Nadu, India.
E-mail: pranabjyoti333@rediffmail.com
Ph: 0091 9442181442.

ABSTRACT: The earlier inventories of the mosquito fauna from Nilgiri hills are compiled and updated the faunal record. Out of 333 species belonging to 47 genera documented in India, 119 species of 21 genera are recorded from Nilgiri hills. 19 species of 4 genera having medical importance are being recorded from Nilgiri hills alone out of total 28 species belong to 5 genera recorded in India. Latter on during the course of dengue vector surveillance in between 2010 – 12 we have added two more species to the check list , An. Stephensi an area distribution record and Toxorhynchites species new under description. (Unpublished data, National Centre for Disease Control, Coonoor Branch). Overall 119 species belong to 21 genera are included in the check list. Evidently the area having richest mosquito fauna contributing more than 35% of the total Indian faunal record of which nineteen species belonging to four genera are considered to be having medical importance recorded from Nilgiri hills. Evidently the area having richest mosquito fauna contributing more than 35% of the total Indian faunal record which includes 19 species (>60%) which are considered having medical importance. The bionomics of the prevailing vectors is discussed.

KEYWORDS: Species, Prevailing, Medical importance, An. stephensi, Toxorhynchites.

INTRODUCTION: There are about 3,500 species of mosquitoes world over globally, ranging in distribution from Sea Level to 3,600 meters altitudes. In India over 330 species including subspecies and varieties are catalogued and yet there may be more number of taxa awaiting description (Knight and Stone 1 Tewari et al 2. Of these about 28 species belonging to five genera are considered to be having medical importance like vectors / secondary vectors of malaria, lymphatic filariasis, Japanese encephalitis, dengue and chickungunya. (Table– 1). In order to have firsthand and correct information on the mosquito fauna it is considered inevitably indispensable to enumerate the biodiversity of mosquito in a given area and there is a need to update the faunal record. Earlier couple of inventories of the Indian mosquito fauna was compiled by Christophers 3 and Barraud 4, and they have recorded 28 species with certainty from Nilgiri hills. Subsequently, studies were carried out by Russel and Jacob 5, Wattal etal 6, Kalra and Wattal 7, Tewari etal 2, 8 and they have brought on record of an additional 89 species including 2 new species description Culex (Lophoceraomyia) raghavanii Rahman, Chowdhury and Kalra , Tewarius reubenae Tewari and Hiriyan 2, 9. Latter on during the course of dengue

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vector surveillance in between 2010 – 12 we have added two more species to the check list i.e., *An. Stephensi* an area distribution record and *Toxorhynchites* species new under description. (Unpublished data, National Centre for Disease Control, Coonoor Branch). Overall 119 species belong to 21 genera are included in the check list. Of these nineteen species belonging to four genera which are considered to be having medical importance are recorded from Nilgiri hills. The bionomics of the above vector species is discussed.

The objective of the study is to update the checklist of mosquito fauna with vector bionomics in the Nilgiris, Tamil Nadu.

STUDY AREA: The Nilgiri hills (Fig-1) bordering Kerala, Karnataka and Tamil Nadu states in the part of Western Ghats lie between Latitude 11° 10' N and Longitude 76° 51' E and situated at altitude ranging from 200 to 2600 Meters above Sea Level. The area receives good rain from both Southwest (July – September) and Northeast (October – December) monsoons. The prevailing average temperature is 5-28°C. The area having diverse terrain feature with distinct ecological characteristic and falls under deciduous wet forest type. The vegetation consists of tropical thorny and mixed deciduous forest with only a few patches of subtropical evergreen forest. Afforestation, plantation of Teak, Eucalyptus, Acacia, and Pinus species are observed on the hill ranges. Extensive tea and coffee plantations are also seen present. The climatic conditions like temperature, humidity, rainfall and also the forest cover with diverse breeding habitats render the area highly suitable for mosquito breeding.

This study was carried out with the objective to update the checklist of mosquito fauna with vector bionomics. The earlier records of mosquito fauna from Nilgiri hills from 1933 to 1992 of various workers are compiled along with addition of our present records (NCDC unpublished data). In the course of vector surveillance carried out by NCDC, Coonoor branch during post monsoon of 2010-12, the probable breeding habitats of the dengue vectors around the human habitations in urban and semi urban situations were examined and collected the immature stage of mosquito, brought to the laboratory, reared to adult and identified the species. Identification of the species was following in principal the keys of Christophers 3 and Barraud 4. To keep confirmative with the most recent and statutory amendment on the classification and nomenclature status of genera, subgenera and species recommendation of Harback and Kitching 10 Harback 11, Reinert 12, Reinert et al 13, and Reinert 14 have been followed.

LIST OF MOSQUITO SPECIES RECORDED FROM NILGIRI HILLS:

1. *Anopheles (Anopheles) aitkenii* James.
2. *Anopheles (Anopheles) barbirostris*, Van der Wulp.
3. *Anopheles (Anopheles) barbumbrosus* Strickland and Chowdhury.
4. *Anopheles (Anopheles) crawfordi* Reid.
5. *Anopheles (Anopheles) gigas* Giles.
6. *Anopheles (Anopheles) insulaeflorum* (Swellengrebel and Swellengrebel de Graaf.)
7. *Anopheles (Anopheles) nigerrimus* Giles.
8. *Anopheles (Anopheles) nilgiricus* Christophers.
9. *Anopheles (Anopheles) peditaeniatus* (Leicester).
10. *Anopheles (Cellia) aconitus* Doenitz.
11. *Anopheles (Cellia) annularis* Van der Wulp.
12. *Anopheles (Cellia) culicifacies* . Giles.
13. *Anopheles (Cellia) elegans* (James).

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14. *Anopheles (Cellia) fluviatilis* . James.
15. *Anopheles (Cellia) Jamesii* Theobald.
16. *Anopheles (Cellia) jeyporiensis* James.
17. *Anopheles (Cellia) karwari* (James).
18. *Anopheles (Cellia) maculates* Theobald.
19. *Anopheles (Cellia) majidi* Young and Majid.
20. *Anopheles (Cellia) mirans* . Sallum and Peyton.
21. *Anopheles (Cellia) moghulensis* Christophers.
22. *Anopheles (Cellia) pallidus* Theobald.
23. *Anopheles (Cellia) splendidus koidzumi*.
24. *Anopheles (Cellia) stephensi* Liston.
25. *Anopheles (Cellia) subpictus* Grassi.
26. *Anopheles (Cellia) tessellates* Theobald.
27. *Anopheles (Cellia) vagus* Doenitz.
28. *Anopheles (Cellia) varuna* Iyengar.
29. *Anopheles (Cellia) willmori* (James).
30. *Aedes (Aedimorphus) alboscuteclatus* (Theobald).
31. *Aedes (Aedimorphus) caecus* (Theobald).
32. *Aedes (Aedimorphus) jamesi* (Edwards).
33. *Aedes (Aedimorphus) pipersalatus* (Giles).
34. *Armigeres (Armigeres) subalbatus* (Coquilett).
35. *Armigeres (Leicesteria) annularis* Leicester.
36. *Armigeres (Leicesteria) digitatus* (Edwards).
37. *Armigeres (Leicesteria) flavus* (Leicester).
38. *Christophersiomyia annulirostris* (Theobald).
(= *Aedes (Christophersiomyia) annulirostris*)
39. *Culex (Culex) barraudi* Edwards.
40. *Culex (Culex) edwardsi* Barraud.
41. *Culex (Culex) fuscocephala* Theobald.
42. *Culex (Culex) gelidus* Theobald.
43. *Culex (Culex) Jacksoni* Edwards.
44. *Culex (Culex) mimeticus* Noe.
45. *Culex (Culex) mimuloides* Barraud.
46. *Culex (Culex) mimulus* Edwards.
47. *Culex (Culex) murrelli* Lien.
48. *Culex (Culex) nilgircus* Edwards.
49. *Culex (Culex) pseudovishnui* Colless.
50. *Culex (Culex) quinquefasciatus* Say.
51. *Culex (Culex) sitiens* Wiedemann.
52. *Culex (Culex) vishnui* Theobald
53. *Culex (Culex) whitmorei* (Giles).
54. *Culex (Culiciomyia) bailyi* Barraud.
55. *Culex (Culiciomyia) pallidothorax* Theobald.
56. *Culex (Eumelanomyia) brevialpis* (Giles).
57. *Culex (Eumelanomyia) foliates* Brug.
58. *Culex (Eumelanomyia) malayi* (Leicester).

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59. *Culex* (*Eumelanomyia*) *pluvialis* Barraud.
60. *Culex* (*Lophoceraomyia*) *cinctellus* Edwards.
61. *Culex* (*Lophoceraomyia*) *flavicornis* Barraud.
62. *Culex* (*Lophoceraomyia*) *lasiopalpis* Sirivanakarn.
63. *Culex* (*Lophoceraomyia*) *mammilifer* (Leicester).
64. *Culex* (*Lophoceraomyia*) *minor* (Leicester).
65. *Culex* (*Lophoceraomyia*) *raghavanii* Rahman, Chowdhury and Kalra.
66. *Culex* (*Oculeomyia*) *bitaeniorhynchus* Giles.
(= *Culex* (*Culex*) *bitaeniorhynchus*)
67. *Culex* (*Oculeomyia*) *infula* Theobald.
(= *Culex* (*Culex*) *infula*)
68. *Downsiomyia* *albolateralis* (Theobald).
(= *Aedes* (*Finlaya*) *albolateralis*)
69. *Fredwardsius* *vittatus* (Bigot)
(= *Aedes* (*Stegomyia*) *vittatus*)
70. *Heizmannia* (*Heizmannia*) *chandi* Edwards.
71. *Heizmannia* (*Heizmannia*) *greenii* (Theobald).
72. *Heizmannia* (*Heizmannia*) *indica* (Theobald).
73. *Heizmannia* (*Heizmannia*) *metallica?* (Leicester).
74. *Heizmannia* (*Mattinglyia*) *discrepans* (Edwards)
75. *Kenknightua* *dissimilis* (Leicester).
(*Aedes* (*Finlaya*) *dissimilis*)
76. *Lutzia* (*Metalutzia*) *fuscanas* Wiedemann.
(= *Culex* (*Lutzia*) *fuscanas*)
77. *Lutzia* (*Metalutzia*) *halifaxii* Theobald.
(= *Culex* (*Lutzia*) *halifaxii*)
78. *Lutzia* (*Metalutzia*) *raptor* Edwards.
(= *Culex* (*Lutzia*) *raptor*)
79. *Lutzia* (*Metalutzia*) *vorax* Edwards.
(= *Culex* (*Lutzia*) *vorax*)
80. *Malaya* *genurostris* Leicester.
81. *Malaya* *jacobsoni* (Edwards).
82. *Mansonia* (*Mansonioides*) *indiana* Edwards.
83. *Mansonia* (*Mansonioides*) *uniformis* (Theobald).
84. *Ochlerotatus* (*Bruceharrisonius*) *aureostriatus* (Doleschall).
(= *Aedes* (*Finlaya*) *aureostriatus*)
85. *Ochlerotatus* (*Finlaya*) *albotaeniatis* (Leicester).
(= *Aedes* (*Finlaya*) *albotaeniatis*)
86. *Ochlerotatus* (*Finlaya*) *chrysolineatus* (Theobald).
(= *Aedes* (*Finlaya*) *chrysolineatus*)
87. *Ochlerotatus* (*Finlaya*) *gubernatoris* (Giles).
(= *Aedes* (*Finlaya*) *gubernatoris*)
88. *Ochlerotatus* (*Finlaya*) *harveyi* Barraud.
(= *Aedes* (*Finlaya*) *harveyi*)
89. *Ochlerotatus* (*Finlaya*) *inquinatus* Edwards.
(= *Aedes* (*Finlaya*) *inquinatus*)

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90. *Ochlerotatus* (Finlaya) *macdaugalli* Edwards.
(=Aedes (Finlaya) *macdaugalli*)
91. *Ochlerotatus* (Finlaya) *oreophilus* Edwards.
(=Aedes (Finlaya) *oreophilus*)
92. *Ochlerotatus* (Finlaya) *pseudotaeniatus* (Giles).
(=Aedes (Finlaya) *pseudotaeniatus*)
93. *Orthopodomyia* *anopheloides* (Giles).
94. *Orthopodomyia* *flavithorax* Barraud.
95. *Paraedes* *menoni* (Mattingly).
96. *Stegomyia* *aegypti* (Linnaeus).
(=Aedes (Stegomyia) *aegypti*)
97. *Stegomyia* *albopicta* (Skuse).
(=Aedes (Stegomyia) *albopictus*)
98. *Stegomyia* *desmotes* Giles.
(=Aedes (Stegomyia) *desmotes*)
99. *Stegomyia* *krombeini* (Huang).
(=Aedes (Stegomyia) *krombeini*)
100. *Stegomyia* *pseudalbopicta* Borel (=Aedes (Stegomyia) *pseudalbopicta*)
101. *Stegomyia* *w-albus* Theobald, (=Aedes (Stegomyia) *w-albus*)
102. *Tewarius* *reubenae* (Tewari and Hiriyan).
103. *Tripteroides* (*Rachionotomyia*) *affinis* (Edwards).
104. *Tripteroides* (*Rachionotomyia*) *aranoides* (Theobald).
105. *Tripteroides* (*Rachionotomyia*) *serratus* (Barraud).
106. *Uranotaenia* (*Pseudoficalbia*) *bicolor* Leicester.
107. *Uranotaenia* (*Pseudoficalbia*) *luteola* Edwards.
108. *Uranotaenia* (*Pseudoficalbia*) *novobscura* Barraud.
109. *Uranotaenia* (*Pseudoficalbia*) *ohamai* Tanaka, Mizusawa and Saugstad.
110. *Uranotaenia* (*Pseudoficalbia*) *stricklandi* Barraud.
111. *Uranotaenia* (*Uranotaenia*) *annandalei* Barraud.
112. *Uranotaenia* (*Uranotaenia*) *kempestris* Leicester.
113. *Uranotaenia* (*Uranotaenia*) *orientalis* Barraud.
114. *Verrallina* (*Neomacleaya*) *agrestis* (Barraud)
(=Aedes (Aedes) *agrestis*).
115. *Verrallina* (*Neomacleaya*) *cautus* (Barraud)
(=Aedes (Aedes) *cautus*).
116. *Verrallina* (*Neomacleaya*) *pseudomediofasciatus* (Theobald).
(=Aedes (Aedes) *pseudomediofasciatus*)
117. *Toxorhynchites* (*Toxorhynchites*) *minimus* (Theobald).
118. *Toxorhynchites* (*Toxorhynchites*) *splendens* (Wiedemann).
119. *Toxorhynchites* (*Toxorhynchites*) new Species *

- **Toxorhynchites* species new under description. This species closely similar to *Tx.edwardsi* Barraud and *Tx.splendens* (Wiedemann) but differ in key characters. These specimens are deposited in the museum of Centre for Research in Medical Entomology (ICMR), Madurai, India with a request for detailed description.
- Mosquitoes of medical importance in India (Table-1):

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- *Anopheles barbirostris*: Isolations of JE virus have been made from this species in Asansol and Bankura in West Bengal and Batai virus from Brahmapalli in Andhra Pradesh Reuben et al 15
- *Anopheles peditaeniatus*: JE virus had been isolated from *Anopheles hyrcanus* group (which include *An. Peditaeniatus*) in Asansol and Bunkura, West Bangal. Chakravarty et al.,16, Benerji et al.,17 and also JEV was isolated from *An. Peditaeniatus* in Mandiya, Karnataka. Mourya et al.18
- *Anopheles annularis*: This belong to annularis group consisting five species, four of these including *An. annularis* had been incriminated as vector of malaria. Harrison, 19.
- *Anopheles culicifacies*: It is one of the most important malaria vector in rural and semi urban areas in India, being responsible for transmitting about 60-70% of malaria infections.
- *An. Culicifacies* complex having five sibling species (A,B,C,D & E) and members of this complex may differ in biologic characteristics influencing epidemiology and control of malaria Subbarao,20
- *Anopheles fluviatilis*: It is one of the most efficient vectors of malaria in India particularly in foothills (Rao ,21 Das et al 22 , Nagpal and Sharma 23,. It was abundant in Western Ghats ranges during pre DDT era where it was playing the role for hyperendemic malaria Russel and Jacob5, Tewari et al.,8
- *Anopheles mirans*: This species belong to *Anopheles leucosphyrus* group and it was misidentified as *An. elegans* Sallum et al.,24 which was reported as vector of simian malaria from foot hills of Nilgiris.
- *Anopheles stephensi*: It is recognized as a major urban malaria vector Rao 21 and it was also considered as a vector in rural area Neogy and Sen 25,
- *Anopheles subpictus*: JE virus has been isolated from this species in Karnataka, Kerala and Tamil Nadu. George et al.,26 ,Dhanda et al.,27 ; *An. Subpictus* appears to play the role of secondary vector of malaria. Panicker et al.28, and also it has been demonstrated to transmit JEV as a bridge vector. Thenmozhi et al.,29
- *Anopheles varuna*: It has been found infected in nature with sporozoites Christophers 3.
- *Culex fuscocephala*: This species is an efficient vector of JE in Thailand Gould et al.30, . Isolations of JE virus from wild – caught mosquitoes have been made from that country and also in India Amerasinghe et al.,31, Reuben et al.,32.
- *Culex gelidus* : This species is considered to be one of the most important vectors of JE in Sri Lanka, Thailand, Malaysia, Vietnam and Sarawak Gould et al.,33, Macdonald et al.,34, Peiris et al.,35. Relatively few isolates have been made in India Reuben et al.,15.
- *Culex vishnui* Subgroup: Three species of this subgroup viz., *Cx. tritaeniorhynchus*, *Cx. vishnui* and *Cx. pseudovishnui* are vectors of JE in India Reuben et al,32
- *Culex quinquefasciatus*: It is well established as principal vector of nocturnal periodic bancroftian filariasis in South Asian countries including India, Myanmar, Malaya, Indonesia, Vietnam. Ragavan 36, Rajagopalan et al,37, 38, 39
- *Culex whitmorei*: Several isolations of JE virus have been made from this species in India and Sri Lanka where it suspected to play a role as a secondary vector (Carey et al.,40 , Peiris et al.,35 , Philip Samuel et al., 41.
- *Mansonia* Species: The *Mansonia* species are the vectors of *Brugia malayi* filariasis in India. Ragavan 36 . In a recent outbreak of JE in Kerala, isolation of JE virus was made from *Ma.uniformis* and *Ma.indiana* Dhanda et al.,27.

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- *Stegomyia aegypti*: A highly anthropophilic, daytime biting mosquito species and principal vector of dengue in Southeast Asia including India where it was responsible for major epidemics Gubler and Kuno 42, WHO, SEARO 43,. Various strains of dengue and chikungunya viruses have been isolated from this species in southern India (Reuben et al., 15, Tewari et al., 44 including demonstration of vertical transmission of dengue virus Thenmozhi et al., 45 .
- *Stegomyia albopicta* :It is typically rural dengue vector and causing mild or asymptomatic dengue virus infection in humans Haweley 46 . A strain of dengue virus (DEN – 4) was isolated from this species in India Reuben et al.,15 and various strains of dengue, Reovirus – 3 and Tembusu virus have been isolated from Southeast Asian countries Huang 47 . Dengue virus (DEN – 2) was isolated from rural areas of Vellore district in South India where it was considered as a secondary vector Tewari et al.,44. In Kerala (South western India) it was abundantly found biting man in outdoors near human habitation. Recently resurgent of dengue reported where DEN 2 was isolated from this species Tyagi et al.,48.

RECENT RECORDS FROM NILGIRIS: The Nilgiri hills having the richest have a rich mosquito fauna contributing over 35% of the total Indian fauna along with the prevalence of 19 species which are considered to be having medical importance and yet the hill ranges are free from any of the mosquito borne diseases. However, once Gudalur taluk was considered as malaria endemic area and *Anopheles fluviatilis* was responsible for the transmission of malaria. After the DDT era during 1940s malaria incidence was declined and the vector *Anopheles fluviatilis* is considered almost vanished extinct species thereafter. The other malaria vectors *Anopheles culicifaciens* and *Anopheles stephensi* are prevailing prevalent in very low density. Tewari et al.,8. *An. Stephensi* an area distribution record as so far it has not been recorded from Nilgiri hills and this is the first report from high altitude area above 2400 MSL, Coonoor. Presently most of the vectors or the species with epidemiological implication are prevailing in very low density. The reason of non endemicity can be attributed to the prevailing ecological conditions such as very low density of vectors, the biologic characteristic of the vector with little influencing epidemiology, low temperature (The area experiencing the average temperature range is 5-25°C.), lesser man mosquito contact. (Most of the mosquito ceases to bite below do not bite at temperatures below 16°C. It is also understood from laboratory based study on vector biology that delimiting factors of dengue incidence i.e., poor percentage of egg hatchability, prolonged period in larval development, poor survival rate in larval stages. All these can be considered a common factor for delimiting the mosquito borne diseases. However *Stegomyia albopicta* which is the dominant species, wide spread throughout the hill ranges accounting for over 65% of the total mosquito specimens recorded. (Unpublished data, NCDC, Coonoor branch). The surrounding areas of the Nilgiri hills particularly parts of Kerala and Tamil Nadu states are often experiencing with dengue infection and *Stegomyia albopicta* is considered the lone epidemic vector of dengue in Kerala state. *Stegomyia albopicta* has the ability to propagate and establish its population displacing *Stegomyia aegypti*, the domesticated mosquito from its domain. Evidently it has been observed that *Stegomyia albopicta* is now fast approaching its establishment nearer to human habitation in peri-urban and urban situations in Nilgiris. (Unpublished data, NCDC, Coonoor branch). Nilgiri hills due to its tourist importance having a large number of floating population from all over the country. When the suitable ecological conditions are in place with susceptible human population and vector abundance along with the

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presence of both vector species, favour the probable increased risk of dengue outbreaks. Hence a constant vigil needs to be undertaken to monitor the dengue incidence in Nilgiri hills. Though it is unpublished data but presented these findings in the 6th Conference of Medical Arthropodology organized by the CRME, (ICMR), Madurai, (TN), from 18th & 19th, October, 2012 & the 57th Annual Conference of IPHA, 2013 at Kolkata from 1st to 3rd Feb, 2013.

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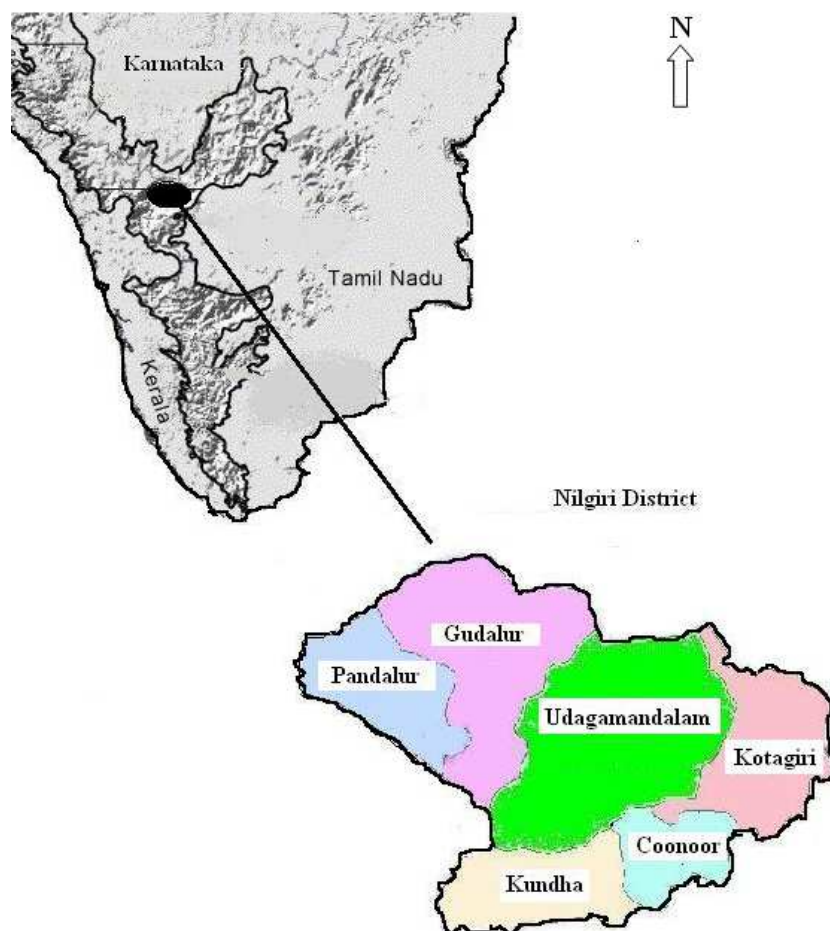
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Table: 1- LIST OF MOSQUITO SPECIES OF MEDICAL IMPORTANCE IN INDIA.

Vectors	Diseases
Anopheles barbirostris *	Malaria, Japanese encephalitis
Anopheles peditaeniatus *	Japanese encephalitis
Anopheles annularis *	Malaria
Anopheles culicifacies *	Malaria
Anopheles fluviatilis *	Malaria
Anopheles minimus	Malaria
Anopheles mirans *	Simian malaria
Anopheles philippinensis	Malaria
Anopheles stephensi *	Malaria
Anopheles subpictus *	Malaria, Japanese encephalitis
Anopheles sondaicus	Malaria
Anopheles varuna *	Malaria
Culex fuscocephala *	Japanese encephalitis
Culex gelidus *	Japanese encephalitis
Culex pseudovishnui *	Japanese encephalitis
Culex quinquefasciatus *	Wuchereria bancrofti filariasis
Culex tritaeniorhynchus	Japanese encephalitis
Culex vishnui *	Japanese encephalitis
Culex whitmorei *	Japanese encephalitis
Culex bitaeniorhynchus	Japanese encephalitis
Culex epidesmus	Japanese encephalitis
Culex infula	Japanese encephalitis
Mansonia annulefera	Brugia malayi filariasis
Mansonia Indiana *	Japanese encephalitis
Mansonia uniformis *	Brugia malayi filariasis , Japanese encephalitis
Stegomyia aegypti *	Dengue, Chikungunya
Stegomyia albopicta *	Dengue, Chikungunya
Downsiomyia nivea	Wuchereria bancrofti filariasis

Fig:1 Map showing study area in Western Ghats



1. Map showing study area in Western Ghats.

2. Map of the Nilgiri district